Denosumab is an anti-bone resorptive drug consisting of complete human monoclonal antibodies that targets receptor activator of nuclear factor B ligand (RANKL), which is responsible for osteoclast formation. The drug has been adapted for bone diseases, such as osteoporosis and bone metastasis related to cancer, but is not used for alveolar bone destruction related to periodontitis. In the present study, we aimed to clarify whether denosumab prevents bone destruction associated with lipopolysaccharide (LPS)-induced calvaria inflammation and experimental periodontitis in model mice. Denosumab does not bind to mouse RANKL, thus we used anti-mouse monoclonal RANKL antibodies. We also examined the inhibitory effects toward bone destruction of another anti-bone resorptive drug zoledronate, a nitrogen-containing bisphosphonate. Local administration of anti-RANKL antibodies into the calvaria area inhibited LPS-induced osteoclast formation and bone destruction, while zoledronate inhibited bone destruction but not osteoclast formation due to its different action mechanism. In periodontitis model mice, in which the second molars were ligated with a silk suture to induce inflammation, intraperitoneal administration of anti-RANKL antibodies significantly inhibited alveolar bone destruction and tooth root exposure. On the other hand, zoledronate only weakly repressed alveolar bone destruction and failed to inhibit root exposure. These results suggest that denosumab is a promising candidate to prevent alveolar bone destruction associated with periodontitis.
periodontitis model was somehow different from that of osteoporosis, thus resulting in our findings regarding the action of zoledronate.

2) How do you think possible side effects of Denosumab? For example, whether its local administration can affect on other bones?
Systematic administration with an intraperitoneal injection of zoledronate as well as that with anti-RANKL antibodies significantly increased bone mass in the femurs of the periodontitis model mice, whereas local injection to calvaria did not have such effects (Supplementary Fig. 1).

In response to questions 1) and 2) above, we have added the following to the Discussion section of the revised manuscript.
“The reason why zoledronate did not inhibit alveolar bone as well as the anti-mouse RANKL antibodies is not clear. We speculate that the microenvironmental condition of alveolar bone destruction in periodontitis model mice was somehow different as compared to that of osteoporosis, thus resulting in our findings for the action of zoledronate. Systematic administration with intraperitoneal injection of zoledronate or anti-RANKL antibodies significantly increased femur bone mass in the periodontitis model, whereas neither given as a local injection showed such an effect (Supplementary Fig. 1). Pharmacokinetic monitoring of bones and femurs affected by periodontitis for comparisons of zoledronate and anti-RANKL antibodies is necessary to reveal the different action mechanisms of these drugs.”

3) I couldn’t find the description about the administration procedure of anti-mouse monoclonal RANKL antibodies in experimental periodontitis. Please indicate it in the materials and methods section.
We apologize that the procedure for administration of the drugs in the periodontitis model mice was not described in the original version of the text. In the revised version, we have modified that portion to provide a description of the procedure used, as follows.
“Under anesthesia with isoflurane inhalation, the cervix of the maxillary left second molar in mice was ligated with a silk suture (size 5-0, MATSUDAIIKA KOGYO Co., Ltd., Tokyo, Japan) 14), then an intraperitoneal administration of saline (200 µl), anti-RANKL antibodies (3 mg/kg), or zoledronate (0.2 mg/kg) was given. At 0, 1, and 2 weeks after ligation and drug administration, each mouse head was subjected to µCT scanning under anesthesia.”

4) How do you think whether Denosumab can be effective if administration of Denosumab starts after partial development of periodontitis? Please indicate your thoughts in the discussion section.
As suggested by the reviewer, we agree that it is important to use denosumab for periodontitis treatment. We think that an administration of anti-RANKL antibodies interrupts the progress of alveolar bone destruction. However, whether alveolar bone volume increases as a result or not is unclear. Regarding the reviewer’s question, we have added the following to the revised Discussion section.
“The present study shows results of pre-treatment with anti-RANKL antibodies in periodontitis model mice, whereas post-treatment effects were not examined. We speculate that anti-RANKL antibodies interrupt destruction and stop the decrease in alveolar bone volume. On the other hand, a dramatic increase in alveolar bone volume is not anticipated when the regenerative potency of alveolar bone is reduced in patients. Additional analyses are necessary to reveal the precise effects of denosumab on periodontal bone disease.”

Additional Information:

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| Method section of manuscript mentions the name of the approving institutional review board and/or ethics committee(s). as follow-up to 
  [Animal Research ] | Yes |
| Your manuscript deals with scientific investigations involving animal studies. |        |
| [Natural Products and Crude Extract Materials (NP/CEM) or not]         | No     |
| Your manuscript deals with NP/CEM.                                    |        |
| Has this manuscript been carefully reviewed by an experienced scientist whose first language is English? | Yes    |
| Name of the English checker [company] or [person + country of his/her first language (mother tongue)] | [Intermed English Services] |
January 23, 2018

Professor Sumio Ohtsuki
Editor-in-Chief
Biological and Pharmaceutical Bulletin

Dear Dr. Ohtsuki,

We are grateful for the reviewer’s evaluation of our manuscript entitled “Anti-mouse RANKL Antibodies Inhibit Alveolar Bone Destruction in Periodontitis Model Mice” (MS18-00026) and providing the excellent suggestions. We have addressed all of the comments from the reviewer, which were very helpful to improve our study, and our revised manuscript is enclosed for your consideration.

In response to those comments and suggestions, we have added text as well as a supplementary figure, and we believe that the revised version adequately addresses the points raised. On behalf of all of the authors, we thank you once again for considering our manuscript for publication in Biological and Pharmaceutical Bulletin, and look forward to hearing from you.

Sincerely yours,

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Anti-mouse RANKL antibodies inhibit alveolar bone destruction in periodontitis model mice

Administration

<table>
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<th>Saline (Control)</th>
<th>Anti-mRANKL antibodies</th>
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<tr>
<td>Destruction of alveolar bone</td>
<td>Inhibited destruction of alveolar bone</td>
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<td>Exposed tooth root</td>
<td>Inhibited exposure of tooth root</td>
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The right second molars of mice were ligated with a silk suture to induce periodontitis. After 2 weeks, the oral cavities were analyzed by μCT to construct 3-D images. Note that the alveolar bone of the control mouse (left) is destructed severely by osteoclasts, however, that of the mouse administrated with anti-mouse RANKL antibodies (right) was protected against destruction.
Anti-mouse RANKL Antibodies Inhibit Alveolar Bone Destruction in Periodontitis Model Mice

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M. Kuritani and N. Sakai contributed equally to this study.
ABSTRACT

Denosumab is an anti-bone resorptive drug consisting of complete human monoclonal antibodies that targets receptor activator of nuclear factor κB ligand (RANKL), which is responsible for osteoclast formation. The drug has been adapted for bone diseases, such as osteoporosis and bone metastasis related to cancer, but is not used for alveolar bone destruction related to periodontitis. In the present study, we aimed to clarify whether denosumab prevents bone destruction associated with lipopolysaccharide (LPS)-induced calvaria inflammation and experimental periodontitis in model mice. Denosumab does not bind to mouse RANKL, thus we used anti-mouse monoclonal RANKL antibodies. We also examined the inhibitory effects toward bone destruction of another anti-bone resorptive drug zoledronate, a nitrogen-containing bisphosphonate. Local administration of anti-RANKL antibodies into the calvaria area inhibited LPS-induced osteoclast formation and bone destruction, while zoledronate inhibited bone destruction but not osteoclast formation due to its different action mechanism. In periodontitis model mice, in which the second molars were ligated with a silk suture to induce inflammation, intraperitoneal administration of anti-RANKL antibodies significantly inhibited alveolar bone destruction and tooth root exposure. On the other hand, zoledronate only weakly repressed alveolar bone destruction and failed to inhibit root exposure. These results suggest that denosumab is a promising candidate to prevent alveolar bone destruction associated with periodontitis.

Key words
denosumab, periodontitis, bone, osteoclast, molecularly targeted drug, bisphosphonate
INTRODUCTION

Denosumab is a powerful anti-bone resorptive drug produced for treatment of osteoporosis patients who have a high risk for fracture, as well as development of cancer metastasis and giant cell tumors in bone tissue. The drug consists of human monoclonal antibodies that bind to receptor activator of nuclear factor κB ligand (RANKL), a tumor necrosis factor (TNF)-super family cytokine produced by osteoblasts and stromal cells in bone tissues. RANKL plays roles in signal transduction for osteoclast differentiation and function by binding to its receptor RANK (receptor activator of nuclear factor κB) expressed on mature osteoclasts and their precursors. Thus, denosumab inhibits bone destruction caused by enhanced differentiation and function of osteoclasts by interrupting RANKL and RANK interactions.

Destruction of alveolar bone occurs in patients with severe periodontitis, which is characterized by chronic gingival inflammation. Dental plaque, which contains bacterial constituents such as lipopolysaccharide (LPS), stimulates osteoclast differentiation via production of inflammatory factors including interleukin (IL)-1, TNF-α, and prostaglandins (PGs). These factors enhance differentiation of osteoclasts on alveolar bone in both direct and indirect manners, resulting in tooth loss due to alveolar bone destruction.

Regular brushing is known to be the most effective means of keeping the periodontal area clean and preventing periodontitis. However, handicapped or bedridden individuals find it difficult to use a dental brush by themselves, and tend to have severe periodontitis, leading to tooth loss. Therefore, establishment of alternative methods for prevention of alveolar bone destruction accompanied by periodontitis is
very important. One possible method is local administration of an anti-bone resorptive
drug such as denosumab in alveolar bone areas. However, it is not known if that drug is
effective for treatment and prevention of alveolar bone destruction caused by
periodontitis.

In the present study, to clarify whether drug therapy using denosumab can prevent
bone destruction in association with periodontitis, we examined the effects of
anti-mouse RANKL monoclonal antibodies, used to mimic denosumab, on LPS-induced
calvarial bone destruction and alveolar bone destruction related to periodontitis induced
experimentally in mice. In addition, we examined the effects of the bisphosphonate
zoledronate, another type of anti-bone resorptive drug, and compared them with those of
the anti-mouse RANKL monoclonal antibodies in mouse models of bone destruction.

Our results showed that administration of anti-mouse RANKL monoclonal
antibodies strongly inhibited calvarial and alveolar bone destruction, suggesting that
denosumab may be a good candidate drug for treatment of periodontitis-induced
alveolar bone destruction.

MATERIALS AND METHODS

Reagents

Lipopolysaccharide (LPS; Escherichia coli 026:B6, #L2654) was purchased from
SIGMA-Aldrich Co., Ltd. (St. Louis, MO, USA), anti-mouse monoclonal RANKL
antibodies (OYC1®) from ORIENTAL YEAST Co., Ltd. (Tokyo, Japan), zoledronate
(ZOMETA®) from Novartis Pharma K.K. (Basel, Switzerland), physiological saline
from Otsuka Pharmaceutical Co., Ltd. (Tokyo, Japan), and type I collagen gel
(Cellmatrix type I A) from Nitta Gelatin Inc. (Osaka, Japan).

Animals and ethical approval

Eight-week-old C57BL/6j male mice were purchased from Sankyo Labo Service Corporation, Inc. (Tokyo, Japan). All animal experiments were approved by the Showa University Animal Care and Use Committee (approval number: 17041), and conducted according to the ethical guidelines of that institution.

LPS-induced calvarial bone destruction model mice

Mice were anesthetized with isoflurane inhalation, then collagen gel (200 µl) containing LPS (25 mg/kg) with or without anti-RANKL antibodies (3 mg/kg), zoledronate (0.2 mg/kg), or saline (control) was injected into the vertex of the head. After 5 days, calvarial portions were excised and fixed in 70% ethanol for 1 hour. Subsequently, the specimens were examined using micro-computed tomography (µCT) to analyze calvarial structures as well as histomorphological analysis to detect osteoclasts. The most effective dosages of LPS, anti-RANKL antibodies, and zoledronate administered to the mice were selected on the basis of previously reported findings.

Detection of osteoclasts in calvaria

Calvarial specimens excised from mice were fixed in 70% ethanol for tartrate-resistant acid phosphatase (TRAP; a marker of osteoclasts) staining using a conventional method with Naphtol AS-MX phosphate (Sigma-Aldrich) and Fast red (Sigma-Aldrich) dissolved in 0.1 M acetic buffer (pH 5.0) containing 1% tartrate acid...
(Wako Pure Chemical Industries, Ltd., Tokyo, Japan)\textsuperscript{13}. Fixed specimens were embedded in O.C.T. compound (Sakura Finetek Japan Co., Ltd., Tokyo, Japan) and cut into coronal sections, which were stained for TRAP\textsuperscript{13}.

**Experimental periodontitis model**

Under anesthesia with isoflurane inhalation, the cervix of the maxillary left second molar in mice was ligated with a silk suture (size 5-0, MATSUDAÏKA KOGYO Co., Ltd., Tokyo, Japan)\textsuperscript{14}, then an intraperitoneal administration of saline (200 µl), anti-RANKL antibodies (3 mg/kg), or zoledronate (0.2 mg/kg) was given. At 0, 1, and 2 weeks after ligation and drug administration, each mouse head was subjected to \(\mu\)CT scanning under anesthesia. The ligatures in all mice remained in place throughout the experimental period. The exposed length of the tooth root was calculated by comparing \(\mu\)CT images obtained at 0 and 2 weeks.

**\(\mu\)CT analysis**

Three-dimensional digital images of the calvaria in LPS-induced bone destruction model mice were reconstructed from \(\mu\)CT analysis findings using a ScanXmate-L090H (Comscantecno, Co., Ltd., Yokohama, Japan). Histomorphological analysis of the \(\mu\)CT images was performed using a TRI/3D-Bon\textsuperscript{®} system (RATOC System Engineering Co., Ltd., Tokyo, Japan). Three-dimensional digital images of alveolar bones and teeth in periodontitis model mice were reconstructed from \(\mu\)CT analysis findings using an \textit{in vivo} 3D \(\mu\)CT system (R_mCT2, Rigaku Co., Ltd., Tokyo, Japan).

**Statistical analysis**
Statistical analyses were performed using the IBM PASW® statistical software package, version 18.0 (IBM, Chicago IL, USA). Values are presented as the mean ± SD. A p value <0.05 was considered to be significant. For experiments involving 6 factors, data were analyzed using one-way ANOVA or Student's t-test.

RESULTS

Administration of anti-mouse RANKL antibodies but not zoledronate inhibits osteoclast formation induced by LPS in calvaria

Denosumab does not bind to mouse RANKL, thus we used anti-mouse RANKL antibodies in this study. Initially, we examined the effects of anti-mouse RANKL antibodies on LPS-induced osteoclast formation in calvarial specimens. Following injection of LPS with saline, an increase in size of TRAP-positive areas, indicating osteoclast formation, was found along calvaria sutures as compared to the control mice (Fig. 1A, 1B, left upper and lower panels). In contrast, simultaneous injection with anti-mouse RANKL antibodies resulted in dramatically smaller TRAP-positive areas induced by LPS (Fig. 1A, 1B, lower middle panels). In addition, administration of anti-mouse RANKL antibodies reduced the size of the primary TRAP-positive areas in the control mice (Fig. 1A, 1B, upper middle panels). On the other hand, zoledronate failed to inhibit the increase in size of TRAP-positive areas in the LPS-injected and control mice (Fig. 1A, 1B, right upper and lower panels). These results indicate that anti-mouse RANKL antibodies inhibit osteoclast formation induced by LPS in the calvaria, while zoledronate does not have such an effect.
**Anti-mouse RANKL antibodies and zoledronate inhibit bone resorption induced by LPS in calvaria**

Next, we examined the effects of anti-mouse RANKL antibodies and zoledronate on calvarial bone destruction induced by LPS (Fig. 2A, B, C). The square in Figure 2A encloses the portion of the calvaria where the number of resorption pits formed by osteoclasts was counted. The number of resorption pits in LPS-injected mice was significantly increased (Fig. 2B left lower panel, 2C) as compared to the control mice, while injection of anti-mouse RANKL antibodies or zoledronate with LPS resulted in markedly inhibited resorption pit formation (Fig. 2B middle and right lower panels, 2C). These results indicate that both anti-mouse RANKL antibodies and zoledronate inhibit calvarial bone destruction induced by LPS.

**Anti-mouse RANKL antibodies inhibit alveolar bone destruction more effectively than zoledronate in periodontitis model mice**

To examine the effects of anti-mouse RANKL antibodies and zoledronate on alveolar bone destruction caused by periodontitis, we performed intraperitoneal administrations of those agents in mice following ligation of the upper-right second molar with a silk suture to induce periodontal inflammation (Fig. 3A, B, C). Figure 3B shows representative μCT images of upper jaws of periodontitis model mice at 0, 1, and 2 weeks after silk suture ligation of the right molars. After 1 and 2 weeks, apparent loss of alveolar bone was observed around the right but not the left molars in the control mice, which had received an injection of saline (Fig. 3B, left lane panels, 3C). On the other hand, alveolar bone loss in mice that received an injection of anti-mouse RANKL antibodies was significantly inhibited (Fig. 3B, middle lane panels, 3C). In the
zoledronate-injected mice, partial inhibition of alveolar bone loss was observed (Fig. 3B, right lane panels, 3C).

Figure 4 shows representative μCT images of buccal side right molars and alveolar bone samples from periodontitis-model mice. In the control mice that received an injection of saline only, molar root exposure due to alveolar bone destruction occurred within 1 week after silk suture ligation of the right molar (Fig. 4A left lane panels, 4B, 4C). Two weeks later, alveolar bone destruction had stopped, though molar root exposure remained (Fig. 4A left lane panels, 4B, 4C). On the other hand, administration of anti-mouse RANKL antibodies significantly inhibited alveolar bone destruction and exposure of the molar roots as compared to the controls (Fig. 4A middle lane panels, 4B, 4C). The inhibitory effects of zoledronate on alveolar bone destruction was not significantly different as compared to the control, though a tendency for suppression of alveolar bone destruction was observed (Fig. 4A right lane panels, 4B, 4C). Together, these results suggest that anti-mouse RANKL antibodies protect against destruction of alveolar bone associated with periodontitis in a more effective manner than zoledronate.

DISCUSSION

Application of minocycline, an antimicrobial drug, to periodontal pockets is generally performed to treat periodontitis. However, such antimicrobial agents do not show inhibitory effects toward alveolar bone destruction in patients with severe periodontitis. Thus, novel drugs to prevent alveolar bone destruction related to periodontitis are anticipated. In the present study, we found that administration of anti-RANKL antibodies strongly suppressed calvarial bone destruction induced by LPS.
and alveolar bone destruction associated with periodontitis in model mice. Our results suggest that denosumab would be applicable for prevention of alveolar bone destruction seen in periodontitis patients.

LPS, a major constituent of bacteria, induces inflammatory cytokines and prostaglandins via Toll-like receptors, and those inflammatory factors have been suggested to induce alveolar bone destruction associated with periodontitis by enhancing osteoclast differentiation \(^{16,17}\). In the present LPS-induced calvarial bone destruction model, local administration of anti-mouse RANKL antibodies into the calvaria region reduced the number of osteoclasts and suppressed resorption pit formation, indicating that even though production of inflammatory factors was induced by LPS, RANKL signaling is essential to induce osteoclast formation. In contrast, zoledronate failed to reduce the number of osteoclasts induced by LPS, suggesting that its action is independent of RANKL signaling. Weinstein et al. \(^{18}\) reported that the number of giant osteoclasts without bone resorption activity was increased in osteoporosis patients who had been administrated alendronate for a long period and those findings support our results showing that administration of anti-RANKL antibodies, but not zoledronate, can inhibit osteoclast differentiation induced by LPS.

In the present experimental periodontitis model mice, severe alveolar bone destruction was found around the molars within 1 week after ligation with a silk suture, which resulted in tooth root exposure. Although administration of anti-RANKL antibodies as well as zoledronate inhibited alveolar bone destruction, the inhibitory effect shown by zoledronate was lower. Similarly, anti-RANKL antibodies but not zoledronate significantly inhibited tooth root exposure. Therefore, we consider that denosumab may be more useful for prevention of alveolar destruction associated with
periodontitis than zoledronate.

The reason why zoledronate did not inhibit alveolar bone as well as the anti-mouse RANKL antibodies is not clear. We speculate that the microenvironmental condition of alveolar bone destruction in periodontitis model mice was somehow different as compared to that of osteoporosis, thus resulting in our findings for the action of zoledronate. Systematic administration with intraperitoneal injection of zoledronate or anti-RANKL antibodies significantly increased femur bone mass in the periodontitis model, whereas neither given as a local injection showed such an effect (Supplementary Fig. 1). Pharmacokinetic monitoring of alveolar bones and femurs affected by periodontitis for comparisons of zoledronate and anti-RANKL antibodies is necessary to reveal the different action mechanisms of these drugs.

Anti-resorptive agent-related osteonecrosis of the jaw (ARONJ) is induced by bisphosphonates and denosumab, and an important clinical problem. In our experiments, osteonecrosis was not observed in the LPS-induced calvarial bone destruction or experimental periodontitis model (data not shown). However, the possibility of ARONJ development induced by such drugs in periodontitis patients cannot be denied. Additional experiments with long-term administration of anti-RANKL antibodies in mice are necessary prior to clinical application. In conclusion, our results suggest that denosumab is a promising drug for prevention of alveolar bone destruction associated with periodontitis.

The present study shows results of pre-treatment with anti-RANKL antibodies in periodontitis model mice, whereas post-treatment effects were not examined. We speculate that anti-RANKL antibodies interrupt destruction and stop the decrease in alveolar bone volume. On the other hand, a dramatic increase in alveolar bone volume is
not anticipated when the regenerative potency of alveolar bone is reduced in patients. Additional analyses are necessary to reveal the precise effects of denosumab on periodontal bone disease.

ACKNOWLEDGMENTS

We thank Dr. Haruka Fukamachi, Dr. Hirobumi Morisaki, and professor Hirotaka Kuwata (Department of Microbiology, School of Dentistry, Showa University) for their technical assistance. This work was supported in part by the Private University Research Branding Project of the Ministry of Education, Culture, Sports, Science and Technology of Japan (MEXT) to Showa University, and the Industry to Support Private Universities Building up Their Foundations of Strategic Research of MEXT (S1411009, S1201014, S0801016) to M.T., Grant-in-Aids for Scientific Research (B) to M. T. (No. 24659830, 26293398) and to T. N.-K. (No. 25293066), and Grant-in-Aids for Scientific Research (C) to N. S. (No. 17K11993), Grant-in-Aid for Challenging Exploratory Research to T. N.-K. (No. 15K15538), and Grant-in-Aid for Young Scientists (B) to A. K. (No. 16K20655) from the Japan Society for the Promotion of Science, as well as grants-in aid from The Science Research Promotion Fund to A. K., The Uehara Memorial Foundation to T. N.-K, The Naito Foundation to T. N.-K, and the Takeda Science Foundation to T. N.-K.

CONFLICT OF INTERESTS

The authors declare no conflicts of interest associated with this manuscript.
REFERENCES


7) Tanabe N, Maeno M, Suzuki N, Fujisaki K. IL-1 a stimulates the formation of


FIGURE LEGENDS

**Fig. 1.** Effects of anti-mouse RANKL antibodies and zoledronate on LPS-induced calvarial bone destruction.

Saline, anti-mouse RANKL antibodies (3 mg/kg), or zoledronate (0.2 mg/kg) with or without LPS (25 mg/kg) were each suspended in type I collagen gel, then injected into the calvaria area. After 5 days, calvaria specimens were excised and stained for TRAP (A). Excised specimens were also cut and the coronal plane stained for TRAP (B). Areas stained in red indicated the existence of osteoclasts. Representative photographs are shown. Bar in A = 5 mm. Bars in B = 0.1 mm.

**Fig. 2.** Effects of anti-mouse RANKL antibodies and zoledronate on resorption pit formation in calvaria induced by LPS.

Saline, anti-mouse RANKL antibodies (3 mg/kg), or zoledronate (0.2 mg/kg) with or without LPS (25 mg/kg) were each suspended in type I collagen gel and injected into the calvaria area. After 5 days, calvaria specimens were excised and analyzed by μCT to construct 3-D images (A, B). The numbers of resorption pits in areas indicated with a yellow frame were counted (A, B, C). Arrows indicate resorption pits (B). Bar in A = 5 mm. Bar in B = 1 mm. Data are shown as the mean ± S.D. (n=6–8 per group). N.S., not significant.

**Fig. 3.** Effects of anti-mouse RANKL antibodies and zoledronate on alveolar bone destruction in periodontitis model mice.

(A) Schema for producing periodontitis model mice, in which the right second
molars were ligated with a silk suture. After 0, 1, and 2 weeks, oral cavities were scanned and analyzed by μCT to construct 3-D images (B). Blue frame in (A) indicates area of oral cavity presented in (B). Alveolar bone volume in the area of the right second molars was analyzed by μCT (C). Data are expressed as the mean ± S.D (n=6). Bar = 1 mm.

**Fig. 4.** Effects of anti-mouse RANKL antibodies and zoledronate on tooth root exposure in periodontitis model mice.

The right second molars in periodontitis model mice were ligated with a silk suture. After 0 (0 w), 1 (1 w), and 2 (2 w) weeks, the oral cavities were scanned and analyzed by μCT to construct 3-D images (A). Yellow arrows indicate tooth roots and red arrows alveolar bone. (B) Formula used to calculate length of tooth roots exposed after 2 weeks. (C) Length of exposed tooth roots at 2 weeks after ligation of second molars. Data are shown as the mean ± S.D. N.S., not significant. Bar = 1 mm.

**Supplementary Fig. 1.** Effects of anti-RANKL antibodies and zoledronate on femur bone mass.

(A) Effects of local administrations of anti-mouse RANKL antibodies (anti-mRANKL ab) and zoledronate on bone mass of femur in calvaria inflammation model mice. Mice were anesthetized with isoflurane inhalation, then collagen gel (200 μl) containing LPS (25 mg/kg) with or without anti-RANKL antibodies (3 mg/kg), zoledronate (0.2 mg/kg), or saline (control) was injected into the vertex of the head. After 5 days, the femurs were excised and fixed in 70% ethanol for 1 hour. Subsequently, bone volume/tissue volume (BV/TV) of distal lesions in the femurs was
evaluated using µCT (n=6).

(B) Effects of systematic administration of anti-mouse RANKL antibodies and zoledronate on femur bone mass in periodontitis model mice. Under anesthesia with isoflurane inhalation, the cervix of the maxillary left second molar in mice was ligated with a silk suture, then saline (200 µl), anti-RANKL antibodies (3 mg/kg), or zoledronate (0.2 mg/kg) were administrated in an intraperitoneal manner. Two weeks after ligation and drug administration, each mouse femur was subjected to µCT scanning and BV/TV was evaluated (n=6).

Note that local administration of anti-mouse RANKL antibodies or zoledronate did not increase BV/TV (A), whereas those given as a systematic administration significantly increased BV/TV (B).
Fig. 3

A

Right ← Left
Medial
Distal

Second molar
Ligation with silk suture

C

Alveolar bone volume in area of right second molar (%)

Saline
Anti-mRANKL ab
Zoledronate

\( p < 0.01 \)

B

Saline
Anti-mRANKL ab
Zoledronate

0 W

1 W

2 W
Fig. 4

A

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<td><img src="Saline_1w.png" alt="Image" /></td>
<td><img src="Anti-mRANKL_ab_1w.png" alt="Image" /></td>
<td><img src="Zoledronate_1w.png" alt="Image" /></td>
</tr>
<tr>
<td>2 w</td>
<td><img src="Saline_2w.png" alt="Image" /></td>
<td><img src="Anti-mRANKL_ab_2w.png" alt="Image" /></td>
<td><img src="Zoledronate_2w.png" alt="Image" /></td>
</tr>
</tbody>
</table>

B

Length (μm) of exposed tooth root = b − a

C

<table>
<thead>
<tr>
<th></th>
<th>Length (μm) of exposed tooth root</th>
</tr>
</thead>
<tbody>
<tr>
<td>Saline</td>
<td><img src="Saline_graph.png" alt="Graph" /></td>
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<tr>
<td>Anti-mRANKL ab</td>
<td><img src="Anti-mRANKL_graph.png" alt="Graph" /></td>
</tr>
<tr>
<td>Zoledronate</td>
<td><img src="Zoledronate_graph.png" alt="Graph" /></td>
</tr>
</tbody>
</table>

P < 0.01

N.S.
Supplementary Fig. 1

A

B

Supplementary materials
Click here to download Supplementary materials
Supplemental_Fig1.TIF